Development of a Robust Screening Method for Pathogenicity of *Colletotrichum* spp. on Strawberry Seedlings Enabling Forward Genetic Studies

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**ABSTRACT**


Generation and screening for nonpathogenic mutants is a popular tool for identifying pathogenicity-related genes. Successful application of this technique for plant fungal pathosystems requires reliable and rapid screening procedures. This study reports on the development of a rapid in vitro bioassay enabling large-scale screening and isolation of nonpathogenic mutants of *Colletotrichum gloeosporioides* and *C. acutatum* on strawberry seedlings. Inoculation was carried out on strawberry seedlings at two different developmental stages: 12-week-old (young) and 15-week-old (older) seedlings. A comparison was made between two inoculation techniques, (i) foliar dip and (ii) root soak, at two incubation temperatures (19 and 25°C). Mortality of young seedlings was observed 4 days after inoculation with both species, reaching 50% within 10 days, using both techniques at 25°C. However, mortality of older seedlings was delayed by 4 days compared with that in the young seedlings when using the root-soak method. Disease development decreased in young and older seedlings at the lower temperature. This method also was reliable in determining pathogenicity of the cucurbit-specific *C. magna* that did not cause disease symptoms on strawberry by either inoculation method. The proposed method enabled screening of more than 980 restriction enzyme-mediated integration mutants resulting in a selection of five reduced-virulence isolates. Initial characterization of some of these mutants revealed large differences in germination and appressorial formation compared with pathogenic isolates.

Additional keyword: anthracnose

Anthracnose has become a major constraint in worldwide strawberry (*Fragaria × ananassa* Duchesne) production. The dominant species causing disease symptoms on strawberry are *Colletotrichum acutatum* J. H. Simmonds, *C. fragariae* Brooks, and *C. gloeosporioides* (Penz.) Penz. & Sacc. in Penz. (teleomorph *Glomerella cingulata* (Stoneman) Spauld. & H. Schrenk) (11,15, 21,26). Anthracnose disease of strawberry affects roots, crowns, petioles, leaves (preferentially immature ones), runners, buds, flowers, and fruit (6,9,10). The ability of these pathogens to attack different plant parts, and the fact that several pathogen species are involved, adds to the complex nature of strawberry anthracnose (16). In Israel, anthracnose is one of the most important diseases affecting strawberry and is caused mainly by *C. acutatum*, but occasionally also by *C. gloeosporioides* (11,13). Infection processes of *Colletotrichum* spp. on strawberry have been well documented in previous studies using light and electron microscopy (4,14). The early stages of invasion, including adhesion to host surface, germination, production of a germ tube, host cuticle penetration via appressorium, and colonization, are essentially the same for all *Colletotrichum* spp. (20,24). Recent ultrastructural studies have shown that *C. acutatum*, although a generalist invader, undergoes a very brief biotrophic stage (less than 12 h) before commencing the extended necrotrophic phase (4). Despite the well-documented infection strategy of *Colletotrichum* spp. on strawberry, the molecular bases of pathogenicity are still very much unknown, and factors important for virulence and aggressiveness of *Colletotrichum* spp. on strawberry deserve further investigation.

To advance our understanding of disease mechanisms, pathogenicity genes can be identified by utilizing a large-scale forward genetic approach for screening for reduced or impaired virulence mutants. In general, the following procedures should be available for this purpose: (i) a random mutagenesis system, (ii) an efficient transformation system, and (iii) a rapid, reliable infection bioassay (29). The restriction enzyme-mediated integration (REMI) technique has been applied successfully to isolate pathogenicity-related genes in several plant-pathogenic fungi (2,3,19,29,31,33) and may serve as an insertional mutagenesis method. An efficient transformation system was developed for *C. gloeosporioides* (25) and in our laboratory for *C. acutatum* (14) by electroporation of germinating conidia, which provided an easy and efficient way to obtain stable transformants of this fungus. However, in the past, pathogenicity assays of *Colletotrichum* spp. were designed especially for breeding programs for anthracnose resistance and are not suitable for large-scale screening for reduced-pathogenicity mutants. These techniques involve foliar application of conidia by spraying, pipetting, or injection of seedlings or plants, which ultimately were selected for foliage, stolon, and crown resistance (7,28). Such methods require considerable greenhouse space, are time and labor consuming, and carry a high chance of cross contamination during inoculation; thus, they are not appropriate for a large-scale screening procedure for nonpathogenic mutants. Likewise, strawberry fruit inoculation methods have been developed to study the virulence of (22) and resistance to anthracnose fruit rot (8). Similarly, an inoculation technique of detached petioles was developed for assessing resistance of strawberry lines to *C. acutatum* (18). However, these two latter methods, in which detached organs were used, are inappropriate for assessing pathogenicity, because resistance mechanisms in detached tissues may be reduced.

The main objective of this study was to establish a rapid and reliable bioassay for large-scale screening of putative nonpathogenic *Colletotrichum* mutants on strawberry. The effects of incubation temperature, seedling age, inoculation technique, and isolate were evaluated to determine the conditions best suited for this purpose. A second objective was to prove the efficacy of the proposed screening method by iso-
lating reduced-pathogenicity mutants for future studies.

MATERIALS AND METHODS

Fungal isolates and inoculum preparation. C. acutatum isolate Ca 149 (IMI391664) (10) and C. gloeosporioides isolates C.g 318 (IMI391663) and C.g 342 pathogenic on strawberry (13), C. magn isolate L2.5 (IMI391662) pathogenic and an ultraviolet-induced nonpathogenic isolate (n-path) on cucurbits (12) were used as control isolates in the development of the screening procedure. The fungi were maintained in the dark on modified Mathur’s medium (M3S) as previously described (10). To prepare inoculum, co-
idia were washed from 5- to 6-day-old cultures with distilled water and adjusted to a final desired concentration of 10⁴, 10⁵, or 10⁶ per ml with a hemacytometer. Ap-
proximately 980 transgenic isolates of C. acutatum and C. gloeosporioides were generated (see below) and evaluated for pathogenicity. Mutants were maintained on M3S supplemented with hygromycin B at 60 µg/ml (Calbiochem, San Diego, CA).

Propagation of strawberry seedlings. Seed of strawberry cv. Malach, an anthrac-

nose-susceptible strawberry cultivar, were produced from fruit harvested from De-
cember to March from a commercially cultivated field (Tzofit, Central Sharon region, Israel). The seeds were collected after peeling and drying the skins for 72 h at ±25°C. The seeds then were surface-
disinfested and soaked overnight in sterile deionized water (dH₂O). Seeds were ger-
mminated on moist, finely ground peat-ver-
miculite medium (vol/vol; 1:1) in contain-
ers and overhead irrigated with KNO₃ (2 g/liter) soluble fertilizer solution. Contain-
ers were covered with plastic bags to main-
tain 90 to 100% relative humidity and kept at 25 ± 1°C under continuous fluorescent light for 2 weeks. Thereafter, the plastic bags were removed and the germinated strawberry seedlings were transferred to a greenhouse with supplemental lighting (16-h photoperiod) and temperature of 25 ± 1°C. Plants in the greenhouse were over-
hed irrigated each week with a soluble fertilizer (N-P-K 4:2.5:6 + 6% microele-
ments; Haifa Chemical Corp., Haifa, Is-
rael). After 12 weeks, most of the seed-
ings used for inoculation experiments consisted of four to five true leaves.

Inoculation techniques. Two inocula-
tion methods were evaluated for their ef-
fect on subsequent disease development. The methods included a root-soak method, coni-
idal suspensions from isolates Ca 149, C.g 318, and n-path were used separately by inserting seedling roots into 2-ml Eppendorf tubes containing 1.5 ml of conidial suspensions at different concentrations. Seedlings inoculated by foliar dip were inverted and inserted into a conidial sus-
pension of the three control isolates and inoculum was applied to all the seedling parts, above the roots. Seedlings then were inserted into Eppendorf tubes filled with 1.5 ml of sterile dH₂O. Sterile water was used as a control for noninoculated seed-
lings in both methods. To maintain high relative humidity, inoculated and control seedlings were transferred immediately into boxes covered with plastic bags, and incubated for a period of 20 days in envi-
ronmental chambers at 19 or 25°C. In each experiment, 50 seedlings were tested for each control isolate at each temperature. Twenty seedlings treated with sterile dH₂O served as noninoculated controls. Experi-
ments determining the effect of inoculation method on seedling mortality were re-
peated twice for each control isolate (50 replicate seedlings were tested for each treatment).

Seedling age and disease response. The effect of seedling age on disease re-
response was evaluated at two developmen-
tal stages: (i) young, 12-week-old seed-
ings and (ii) older, 15-week-old seedlings. Both sets of seedlings were grown for the first 12 weeks as described above. Older seedlings were transplanted at 12 weeks to polyethylene trays containing peat-ver-
miculite medium (vol/vol; 1:1) and were grown for an additional 3 weeks in a greenhouse with a 16-h photoperiod. Co-
nidial suspensions of isolates Ca 149, C.g 318, and n-path were used separately to inoculate young or older seedlings at 19 and 25 ± 1°C and mortality was recorded over a 3-week period. Experiments exami-
ining the effect of seedling age were re-
peated twice for each control isolate at each incubation temperature (50 replicate 
seedlings were tested for each treatment).

Fungal transformation. Electropora-
tion of germinating conidia was performed essentially as previously described (14). Briefly, isolates Ca 149, C.g 342, and C.g 318 were cultured on solid M3S medium for 5 days. Conidia were collected in pea juice (25), adjusted to 10⁶ conidia/ml, and incubated at 28°C for 4.5 to 5 h to initiate germination. The germinated conidia were collected, washed with cold electropora-
tion buffer (1 mM n-2-hydroxyethlpi-
perazine-2-ethanesulfonic acid [HEPES], 50 mM mannitol, pH 7.5) and concentrated to 10⁶ conidia/ml, and 100-µl aliquots were distributed in cold electroporation cuvettes (Bio-Rad, Hercules, CA). The 5.13-kb pGH-J plasmid (33) was used as a trans-
formation vector. REMI was performed with 5 µg of linearized XbaI or HinIII pGH-J plasmid for each transformation event (performed in separate cuvettes) in the presence of 24 units of the same en-
zyme used to linearize the plasmid, as previously described (33).

Electroporation was performed with a gene pulser (Bio-Rad) operating at 1.4 kV, 800 Ω, and 25 µF. After application of the electric pulse, the conidia were transferred to regeneration (Reg) medium (145.7 g of mannitol, 4 g of yeast extract [Difco Labo-
ratories, Detroit], 1 g of soluble starch, 16 g of agar [Difco Laboratories], and 50 ml of pea juice per liter). After 10 h, a top overlay (2 mm) of water agar with hygro-
mycin B 125 µg/ml (Calbiochem) was added. Transgenic colonies appeared 4 to 5 days after transformation. Stability of transormants was assessed by serial trans-
fer of mycelial plugs of representative REMI transformants onto fresh Reg me-
dium containing or devoid of hygromycin B at 60 µg/ml every 2 weeks for a 2-month period.

Pathogenicity assays for isolation of putative nonpathogenic mutants. Hygro-
mycin-transformed putative nonpathogenic mutant isolates were tested for their ability to induce anthracnose symptoms on straw-
berry seedlings. Primary pathogenicity screens used five anthracnose susceptible ‘Malach’ seedlings per REMI transfor-
mant. Young seedlings with four to five true leaves were inoculated by the foliar-
dip method with 10⁶ conidia/ml of each REMI isolate and inserted into Eppendorf tubes. Seedling mortality was assessed after 7 to 10 days. Transformants that did not cause disease on any of the five plants were subjected to an additional patho-
genicity screening, using 15 seedlings. REMI transformants exhibiting reduced pathogenicity were screened a third time in the same manner as the second test. All experiments included a positive control that consisted of 15 seedlings inoculated with wild-type isolates of C. gloeo-
sporioides, C.g 318, and C.g 342, and C. acutatum, Ca 149. Negative controls consis-
ted of 15 replicate seedlings inoculated with n-path of C. magn and a water con-
trol. Pathogenicity of the putative mutants was verified further on mature daughter plants. Mature plants were inoculated by foliar dip or sprayed with conidial suspensions of 10⁶ conidia/ml and were main-
tained under 90 to 100% relative humidity by covering with plastic bags at 25 ± 1°C for 2 weeks. Disease severity of daughter plants was assessed after 2 weeks accord-
ing to the following disease severity rating (DSR) scale: 0 = plants with no visible lesions; 1 = one small (<0.5 cm) lesion on leaves or petioles; 2 = two small lesions on leaves or petioles; 3 = one or more large, actively developing lesions on leaves or petioles; 4 = developing necrotic and wilt 
symptoms; and 5 = dead plant with ne-
crotic lesions. Pathogenicity of each poten-
tial mutant and wild-type isolates was expressed as the mean DSR of 10 mature strawberry plants per isolate.

Germination and appressorial forma-
tion. Germination and appressorial forma-
tion by Colletotrichum mutants impaired in pathogenicity and comparative wild-type isolates were observed on GLASTIC SLIDES (HYCOR; Biomedical Inc., Gar-
den Grove, CA). Briefly, 200 μl of a conid-

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ial suspension of 10^5 conidia/ml of each isolate was placed into five cells of the glassic slide and kept on moistened filter paper in a moist chamber. After 12 to 14 h of incubation, conidial germination and appressorial development were observed in three microscopic fields in each of five cells per slide. Three germination and appressorial formation stages were documented: conidial germination and immediate appressorial formation (apressoria), conidial germination and appressorial formation after more than double the conidia length (germination + appressoria), and conidia germination without appressorial formation (conidia + germination). Frequencies of germination and appressorial formation were determined for 400 conidia from each isolate in each of three trials and compared with that of wild-type frequency by a least significance difference test (LSD; P = 0.05). To confirm isolate phenotype, strawberry seedlings were inoculated with conidial droplets and germination and appressorial formation was monitored microscopically after 12 to 14 h on leaves.

Statistical analysis. Data were analyzed using the JMP software package (version 3.2.6; SAS Institute, Inc. Cary, NC). Each in vitro seedling inoculation experiment was repeated once and data from each experimental repeat were subjected to analysis of variance (ANOVA) and error variances were evaluated for homogeneity by pairwise t tests. ANOVA was performed separately for each seedling age × incubation temperature × inoculation method × incubation temperature of each isolate combination. Because there were no significant statistical differences among experiments and the error variances were homogeneous (P > 0.10), data from individual experiments were combined. The combined data were separated using the Tukey-Kramer multiple comparison test, P < 0.05, to detect differences among treatments. Additionally, the time period required to reach 50% seedling mortality was calculated from the respective regression equation of disease progress for each isolate combination. Significant differences of time to reach 50% mortality were calculated by contrast t tests (P < 0.05), in an ANOVA for the linear regression. Frequency of germination and appressorial formation were pooled for each of three replications per isolate after t tests were conducted to determine homogeneity of variance among the three independent experiments, and no significant statistical difference among experiments was found (P > 0.05). Mean comparisons of germination and appressorial formation were calculated with the combined data using LSD, according to the Tukey-Kramer multiple comparison test at P < 0.05.

RESULTS

Disease incidence in strawberry seedlings. Disease incidence and symptom development in the inoculated seedlings were monitored over 20-day periods. After this period, lower leaves closest to the water began to senesce and rot in water and n-path C. magna control treatments. Therefore, the experiments were terminated at this stage. Seeding mortality was determined as a binary test (dead or live seedling) and not by a disease severity rating (DSR) scale, which was used later on mature plants inoculated with putative nonpathogenic mutants. Local anthracnose lesions on leaves and petioles were monitored but seeding mortality was recorded only when all seedling petioles collapsed, which led to seedling death. Disease progress in the seedlings inoculated with the n-path isolate was similar to water controls and was significantly lower than in seedlings inoculated with the pathogenic wild-type isolates (Fig. 1A).

Effect of seedling age on disease response. The effect of seedling age on disease response was evaluated using 12-week-old (young) and 15-week-old (older) seedlings. A rapid disease response, similar for both young and older inoculated seedlings, was observed within 4 days by the foliar dip method, with isolates C.g 318 of C. gloeosporioides and C.a 149 of C. acutatum (Fig. 2A). No significant differences were observed in the time required to reach 50% seedling mortality of young and older seedlings inoculated by the foliar dip method at 25°C (Table 1). However, seeding mortality decreased significantly in older seedlings compared with young seedlings inoculated by the root-soak method, with all pathogenic isolates (Fig. 2B). Mortality of young seedlings was observed from 4 days after root inoculation, whereas that of older seedlings was delayed, occurring 8 days after inoculation (Fig. 2B).

Effect of inoculation method on disease response. Foliar-dip and root-soak inoculation methods were evaluated for their effect on subsequent disease development. Young and older seedlings inoculated by the foliar-dip method developed typical anthracnose lesions on foliage and petioles within 2 to 3 days. However, seedlings inoculated by the root-soak method did not develop lesions on foliage but ex-

Fig. 1. Disease incidence 16 days after inoculation with A, young seedlings inoculated with a non-pathogenic isolate (n-path) of Colletotrichum magn, pathogenic isolate C.g 342 of C. gloeosporioides, and water control, and B, reduced pathogenicity mutants (c-52, g-233, and e-149) and pathogenic mutant (C-17) of C.g 342 of C. gloeosporioides. Seedlings were inoculated by the foliar-dip method in a conidial suspension (10^5 conidia/ml) of each isolate and incubated at 25°C.
hibited active necrotic lesions on lower petioles that resulted in rapid seedling collapse at 25°C. The root-soak technique resulted in rapid seedling collapse but minimal anthracnose symptoms were observed on foliage. No significant differences were observed in time required to reach 50% disease incidence of young seedlings inoculated by the two methods at 25°C (Table 1). In older seedlings, foliar dip produced more rapid disease progress than root soak (Fig. 2A and B) and time required for 50% mortality was significantly less for seedlings inoculated with most isolates by foliar dip than by root soak at 25°C (Table 1).

**Effect of incubation temperature on disease response.** Temperatures of 19 and 25°C were evaluated for their effect on disease development and pathogenicity levels. The onset of seedling mortality occurred 2 days later at 19°C compared with 25°C for both young and older seedlings inoculated by foliar dip (Fig. 2A and C). Significant differences in time required to reach 50% mortality of seedlings were observed only for young seedlings inoculated with C.a 149 by the foliage method at both incubation temperatures (Table 1). Disease progress was much slower for young and older seedlings inoculated at 19°C by the root-soak method with isolate C.g 318 of *C. gloeosporioides* than in young seedlings inoculated with isolate C.a 149 of *C. acutatum* (Fig. 2D).

**Effect of inoculum concentration on disease response.** Inoculum concentrations of $10^4$, $10^5$, and $10^6$ conidia/ml of three isolates C.a 149, C.g 318, and n-path of *C. magna* were assessed for disease response (Table 2). Significant differences in disease progress were observed for isolate C.g 318 of *C. gloeosporioides* at all three inoculum concentrations. Significant differences also were observed for C.g 318 in the time required to reach 50% seedling mortality of 13.8, 9.5, and 6.4 days for $10^4$, $10^5$, and $10^6$ conidia/ml, respectively. Similar trends were observed for isolate C.a 149 of *C. acutatum* at $10^4$ and $10^5$ conidia/ml, corresponding to 10.4, 8.2, and 8.0 days, respectively (Table 2).

**Screening for reduced-pathogenicity mutants.** The foliar-dip method was evaluated for detecting reduced pathogenicity of REMI mutants compared with the n-path mutant of *C. magna*. Significant differences in mortality were observed in seedlings inoculated with the wild-type C.g 342 of *C. gloeosporioides* compared

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**Table 1.** Time required to reach 50% mortality in young (12-week-old) and older (15-week-old) strawberry seedlings by the root-soak and foliar-dip methods at different incubation temperatures with isolate C.a 149 of *Colletotrichum acutatum* and isolate C.g 318 of *C. gloeosporioides*.

<table>
<thead>
<tr>
<th>Isolate</th>
<th>Inoculation method</th>
<th>Incubation temperature (°C)</th>
<th>Days to 50% mortality*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Young</td>
<td>Older</td>
</tr>
<tr>
<td>C.a 149</td>
<td>Foliar dip</td>
<td>25</td>
<td>8.2 a</td>
</tr>
<tr>
<td>C.a 149</td>
<td>Root soak</td>
<td>25</td>
<td>7.7 a</td>
</tr>
<tr>
<td>C.a 149</td>
<td>Foliar dip</td>
<td>19</td>
<td>11.2 b</td>
</tr>
<tr>
<td>C.a 149</td>
<td>Root soak</td>
<td>19</td>
<td>8.2 a</td>
</tr>
<tr>
<td>C.g 318</td>
<td>Foliar dip</td>
<td>25</td>
<td>9.5 ab</td>
</tr>
<tr>
<td>C.g 318</td>
<td>Root soak</td>
<td>25</td>
<td>10.5 b</td>
</tr>
<tr>
<td>C.g 318</td>
<td>Foliar dip</td>
<td>19</td>
<td>9.8 b</td>
</tr>
<tr>
<td>C.g 318</td>
<td>Root soak</td>
<td>19</td>
<td>16.9 c</td>
</tr>
</tbody>
</table>

*Values for 50% seedling mortality were calculated from the respective regression equation of disease progress for each seedling age × inoculation method × isolate × incubation temperature combination. Values with the same letters do not differ significantly ($P < 0.05$) according to contrast $t$ test.

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**Table 2.** Time required to reach 50% mortality in young (12-week-old) and older (15-week-old) strawberry seedlings by the root-soak and foliar-dip methods at different incubation temperatures with isolate C.a 149 of *Colletotrichum acutatum* and isolate C.g 318 of *C. gloeosporioides*.

<table>
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<th>Days to 50% mortality</th>
</tr>
</thead>
<tbody>
<tr>
<td>C.a 149</td>
<td>Foliar dip</td>
<td>25</td>
<td>8.2 a</td>
</tr>
<tr>
<td>C.a 149</td>
<td>Root soak</td>
<td>25</td>
<td>7.7 a</td>
</tr>
<tr>
<td>C.a 149</td>
<td>Foliar dip</td>
<td>19</td>
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<td>C.a 149</td>
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<tr>
<td>C.g 318</td>
<td>Foliar dip</td>
<td>25</td>
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<td>C.g 318</td>
<td>Root soak</td>
<td>19</td>
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</tbody>
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**Fig. 2.** Disease incidence in strawberry seedlings inoculated with wild-type isolates C.g 318 of *Colletotrichum gloeosporioides*, C.a 149 of *C. acutatum*, and nonpathogenic ultraviolet mutant (n-path) of *C. magna*. **A**, Foliar dip of young (Y) and older (O) seedlings at 25°C. **B**, Root-soak inoculation of Y and O seedlings at 25°C. **C**, Foliar dip of Y and O seedlings at 19°C. **D**, Root-soak inoculation of Y and O seedlings at 19°C. Each datum point is the mean of two individual experiments. Distances larger than the vertical bars between symbols at each point are significant ($P < 0.05$) according to Tukey Kramer multiple comparison test.
with REMI mutant isolates c-52, c-63, e-149, and g-233 (Figs. 1B and 3A). Furthermore, wild-type isolate L.2.5 of *C. magna*, pathogenic on cucurbits, did not cause disease in strawberry seedlings (Fig. 3A). REMI mutants that exhibited reduced pathogenicity after three seedling screening tests were used to inoculate mature daughter plants. Disease severity values were significantly lower for plants inoculated with c-52, c-63, e-149, and g-233 compared with wild-type isolate C.g 342 of *C. gloeosporioides* (Table 3).

**Assays for germination and appressorium formation.** Frequency of appressorium formation was determined and compared with that of the wild-type isolates. Significant differences were observed between the n-path mutant compared with that of wild-type L.2.5 isolate of *C. magna*. Significant differences also were observed between the REMI reduced-pathogenicity mutants and the corresponding wild-type C.g 342 isolate of *C. gloeosporioides*. In n-path and REMI mutants c-63, e-149, and g-233, conidia tended to form long germ tubes without appressoria at significantly higher rates than their corresponding wild-type isolates (Fig. 3B). The frequency of conidial germination and initial appressorium formation was significantly higher in wild-type C.g 342 than in reduced-pathogenicity REMI mutants.

**DISCUSSION**

As a first step in evaluating pathogenicity of *Colletotrichum* REMI mutants on strawberry seedlings, it was necessary to develop a rapid, reliable screening protocol. To assess pathogenicity of isolates, it was necessary to clearly define the role of seedling age in susceptibility to anthracnose. Young and older seedlings inoculated by foliar dip were similar in the time required to reach 50% mortality at 25°C (Table 1). Young immature, true leaves are preferentially susceptible to infection by *Colletotrichum* spp. (7,21). Thus, seedling transplanting, which is time and labor consuming, is dispensable for screening pathogenicity by the foliar-dip method. Recent studies have shown plant age to be an important consideration when screening strawberry seedlings for resistance. In general, seedlings become more resistant as they mature (27). In this study, only older seedlings inoculated by the root-soak method seemed to be more resistant at 25°C, compared with young seedlings (Fig. 1B). This may be the result of a more developed root system in older seedlings. Traditionally, older seedlings are used to screen for cultivar resistance to *C. acutatum*, a dip inoculation technique in which entire plants were immersed in a conidial suspension resulted in accurate disease response of susceptible cultivars.

Disease progress was delayed at 19°C compared with 25°C with all pathogenic isolates (Fig. 2), which was in agreement with previous studies showing that a latent infection period decreased as temperatures increased (17,32). Although appearance of disease symptoms was delayed in seedlings inoculated by the leaf method at 19°C, disease incidence progressed and eventually reached levels similar to that at 25°C (Fig. 2C). An incubation temperature of 19°C allowed better discrimination between C.g 318 of *C. gloeosporioides* and

![Graph](image)

**Table 2.** Time required to reach 50% mortality in young strawberry seedlings inoculated by the foliar-dip method with three different inoculum concentrations of isolates C.a 149 of *Colletotrichum acutatum* and C.g 318 of *C. gloeosporioides*.

<table>
<thead>
<tr>
<th>Isolate</th>
<th>Inoculum (conidia/ml)</th>
<th>Time (days)</th>
</tr>
</thead>
<tbody>
<tr>
<td>C.g 318</td>
<td>10⁴</td>
<td>13.8 a</td>
</tr>
<tr>
<td>C.g 318</td>
<td>10³</td>
<td>9.5 b</td>
</tr>
<tr>
<td>C.g 318</td>
<td>10²</td>
<td>6.4 c</td>
</tr>
<tr>
<td>C.a 149</td>
<td>10⁴</td>
<td>10.4 d</td>
</tr>
<tr>
<td>C.a 149</td>
<td>10³</td>
<td>8.2 b</td>
</tr>
<tr>
<td>C.a 149</td>
<td>10²</td>
<td>8.0 b</td>
</tr>
</tbody>
</table>

* Values for 50% seedling mortality were calculated from the regression equation of disease progress for each inoculum concentration and isolate combination. Values with different letters are significantly different based on results of contrast t tests (*P* < 0.05).

![Graph](image)

**Fig. 3.** A, Disease response of strawberry seedlings inoculated with mutant isolates of *Colletotrichum gloeosporioides* and control isolates L.2.5, a nonpathogenic mutant (n-path) of *C. magna*, and C.g 342 of *C. gloeosporioides*. Distances between symbols at each point that are larger than the vertical bars are significant (*P* ≤ 0.05). B, Assessment of conidial germination and appressorial formation compared to wild-type isolates. Each column in B is the mean of three individual experiments. Bars represent standard error of the mean and common letters are not significantly different (*P* ≤ 0.05), according to the Tukey-Kramer multiple comparison test, for each of the parameters (conidia + germination, appressoria) between isolates.
C. a 149 of C. acutatum in seedlings inoculated by the root-soak method (Fig. 2D). Recent studies showed that, at lower temperatures, C. acutatum produced more conidia than C. gloeosporioides (17,23). However, at 25°C, there was little difference in disease incidence among control isolates; therefore, 25°C was chosen as the recommended incubation temperature. Conidial concentration is an important factor in inducing the disease response in strawberry seedlings (Table 2). Significant differences in seedling mortality due to different concentrations of C.g 318 may be explained by lower sporulation of C. gloeosporioides compared with C. acutatum and a shorter latent period of C. acutatum compared with C. gloeosporioides (17). Initial inoculum concentration may have changed over time during incubation and may be important in such a short pathogenicity bioassay. A conidial suspension adjusted to 10^7 conidia/ml was effective in screening for reduced pathogenicity mutants, and well suited for the purpose of these screening tests.

The root-soak and the foliar-dip methods allowed for discrimination of nonspecific host–pathogen interaction, whereby the specific cucurbit pathogen, C. magna, did not cause disease in strawberry seedlings at the conidial concentrations tested (Fig. 3A). In order to screen for reduced pathogenicity mutants, we chose the technique causing typical symptoms on susceptible cultivars that required less time and labor (i.e., foliage inoculation of young seedlings using inoculum concentration adjusted to 10^7 conidia/ml at an incubation temperature of 25°C). This method was consistent for evaluation of pathogenicity of Colletotrichum REMI mutants on strawberry seedlings. The developed technique was reliable and accurate because isolates that were reduced or impaired in pathogenicity on young seedlings during several screenings were reduced consistently or nonpathogenic on plants at all developmental stages, including mature daughter plants (Fig 3A; Table 3). Therefore, this method is recommended for a preliminary screening of a large population of transformants. The proposed method requires considerably less space because Eppendorf tubes are used instead of pots. The rapid screening of potential mutants could not have been performed with such ease with the standard spray inoculation techniques (7,27,28). The foliar-dip method was devised for large-scale screening and isolation of Colletotrichum REMI mutants with reduced pathogenicity. The time required for screening 150 mutants with this procedure, which involves two people, was approximately 8 h.

Five of the REMI mutants impaired in pathogenicity isolated by the rapid technique were characterized for initial parameters of germination and appressorium formation. Formation of the appressorium and penetration of the host are integral parts of the infection process of Colletotrichum spp. (24). In some species, formation of appressoria may be obligatory for infection; whereas, in others, it may be optional or not required at all (35). A mode of germination observed for REMI mutants c-63, e-149, and g-233 consisting of ex- tended germ tube elongation before appressorium formation or the absence of appressorium formation also has been reported for Cochliobolus heterostrophus (5) and Magnaporthe grisea (30). Colletotrichum acutatum shows similar germination behavior on asymptomatic plants (eggplant, pepper, and tomato) which occurred mainly from one side of the conidium and was characterized by long, thin, unbranched hyphae (14). This kind of development occasionally resembles the effect of mutation in signal transduction components observed in other fungal species (1,34). Further characterization of the reduced-pathogenicity mutants will allow us to identify specific genes involved in pathogenicity of Colletotrichum in strawberry.

In this study, a large-scale screening technique was developed for pathogenicity assessment of Colletotrichum isolates and its benefit for selection of reduced-virulence mutants was proven. Application of this procedure may have potential for screening additional pathogens on strawberry, such as Rhizoctonia spp., and for assessing potential biocontrol agents.

**ACKNOWLEDGMENTS**

We thank Y. Nitzani and M. Maymon for technical assistance.

**LITERATURE CITED**


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**Table 3.** Response of mature strawberry daugh- ter plants to inoculation with various restriction enzyme-mediated integration mutants of Colle- totrichum gloeosporioides and C. acutatum reduced in pathogenicity relative to wild-type isolates.

<table>
<thead>
<tr>
<th>Isolates</th>
<th>DSR</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>C.g 342</td>
<td>4.25</td>
<td>a</td>
</tr>
<tr>
<td>N-path</td>
<td>0.00</td>
<td>b</td>
</tr>
<tr>
<td>c-52</td>
<td>0.50</td>
<td>b</td>
</tr>
<tr>
<td>c-63</td>
<td>1.25</td>
<td>d</td>
</tr>
<tr>
<td>e-149</td>
<td>0.75</td>
<td>b</td>
</tr>
<tr>
<td>g-233</td>
<td>0.25</td>
<td>b</td>
</tr>
</tbody>
</table>

* Disease severity rating (DSR) of daughter plants was scored 2 weeks after inoculation for disease symptoms according to a 0-5 scale, where 0 = no visible lesions and 5 = dead plants; values represent the means for 10 plants per isolate. Values with different letters are significantly different at P ≤ 0.05 according to the Tukey-Kramer multiple comparison test.
Germination and sporulation of *Colletotrichum acutatum* on symptomless strawberry leaves. Phytopathology 91:659-664.


